

## Synthesis, *in vitro* antiproliferative and antimycobacterial activity of thiazolidine-2,4-dione and hydantoin derivatives

V.T. Angelova<sup>1\*</sup>, V. Valcheva<sup>2</sup>, N. Vassilev<sup>3</sup>, R. Buyukliev<sup>1</sup>, R. Mihaylova<sup>1</sup>, G. Momekov<sup>1</sup>

<sup>1</sup>Faculty of Pharmacy, Medical University of Sofia, 2 Dunav Str., 1000 Sofia, Bulgaria;

<sup>2</sup>"Stefan Angelov" Institute of Microbiology, Bulgarian Academy of Sciences, 26 Acad. Bonchev Str., 1113 Sofia, Bulgaria;

<sup>3</sup>Institute of Organic Chemistry with Centre of Phytochemistry, Bulgarian Academy of Sciences, 9 Acad. Bonchev Str., 1113 Sofia, Bulgaria

Received February 23, 2017; Revised April 27, 2017

New 2*H*-chromene derivatives bearing thiazolidine-2,4-dione or hydantoin moieties were synthesized and the structures were confirmed by <sup>1</sup>H NMR and <sup>13</sup>C NMR as well as 2D NMR, FTIR and HR-ESI-MS spectra. The compounds were evaluated for their *in vitro* cytotoxicity against four human cancer cell lines, namely HL-60 (acute promyelocyte leukemia), REH (lymphoid leukemia), K-562 (chronic myeloid leukemia) and EJ (urinary bladder carcinoma). The 2*H*-chromene derivative containing thiazolidine-2,4-dione ring **5** was potent against a panel of three cancer cell lines, with an IC<sub>50</sub> in the range of 13.1-39.6 μM and exhibited pronounced antiproliferative activity against lymphoid leukemia (REH cell line) with an IC<sub>50</sub> value of 13.1 μM. The hydantoin containing 2*H*-chromene derivative **7** having an IC<sub>50</sub> value of 83.7 μM was highly effective against chronic myeloid leukemia K-562. Compounds **5** and **7** also demonstrated significant antimycobacterial activity against *Mycobacterium tuberculosis* H37Rv strain with minimum inhibitory concentration (MIC) ranging from 0.29 and 0.36 μM, respectively.

**Keywords:** Antiproliferative/cytotoxic effects, Antimycobacterial activity, 2*H*-Chromene, Hydantoin, Thiazolidine-2,4-dione.

### INTRODUCTION

The hydantoin (1,3-imidazolidinedione) derivatives [1] and thiazolidinediones (thiazolidine-2,4-diones) [2, 3] exhibit a plethora of biochemical and pharmacological activities. The thiazolidinediones (TZDs) have been characterized as a new dawn in cancer chemotherapy with a broad spectrum of cytotoxicity towards different human cancer cells [4, 5]. It is well known that TZDs exert their anti-diabetic effects through a mechanism that involves activation of PPAR $\gamma$  receptor. The wide spectrum of PPAR $\gamma$  activation effects may also be beneficial in the treatment of different types of cancer [6]. Thus, several new drugs such as efatutazone, netoglitazone, rosiglitazone and troglitazone (Fig. 1), exhibit their anticancer activity via PPAR $\gamma$  -dependent or -independent signaling pathways [7]. In the meantime, the antitumor effect of hydantoin derivatives has been reported by a number of authors (Fig. 1). Some of hydantoin derivatives, characterized by a 1-phenethyl and a 5-(*E*)-benzylidene substituent, inhibit EGFR autophosphorylation and polyGAT phosphorylation, as well as inhibit the growth and proliferation of human A431 cells, which

overexpress EGFR [8] (Fig. 1). The hydantoin derivatives tested by Rajic *et al.*, showed rather marked inhibitory activity against HeLa and MCF-7 cell lines, and no cytotoxic effects on normal cells [9]. The anti-cancer potential of 5-benzylidene-hydantoins demonstrating its relation to SIRT inhibition was proved by Lionel *et al.* [10]. Mudit *et al.*, [11] described phenylmethylene hydantoins, as a novel antimetastatic lead class with the potential to control metastatic prostate cancer. The compounds synthesized by Reddy *et al.*, [12] contain an *N*-benzylindole nucleus linked to a hydantoin moiety *via* a double bond with *Z*-geometry were described as potential anticancer agents for the treatment of breast cancer. 5-(1*H*-indol-3-ylmethyl)-2-thiohydantoins and 5-(1*H*-indol-3-ylmethyl)hydantoins were found to be potent necrostatins [13]. In addition, (*Z*)-5-(4-hydroxybenzylidene)-imidazolidine-2,4-dione display moderate antiproliferative activity against the human cervical carcinoma (HeLa) cell line [14].

On the other hand, many studies have shown thatazole heterocycles such as thiazolidine-2,4-dione and imidazolidine-2,4-dione are useful pharmacophores possessing antimycobacterial activity [15]. There have been reports on some thiazolidine-2,4-dione derivatives [16] and various imidazolidine-2,4-dione derivatives [17, 18], evaluated in the primary assay for antimycobacterial activity which exhibit

\* To whom all correspondence should be sent:  
E-mail: violina\_stoyanova@abv.bg;  
violina@pharmfac.net

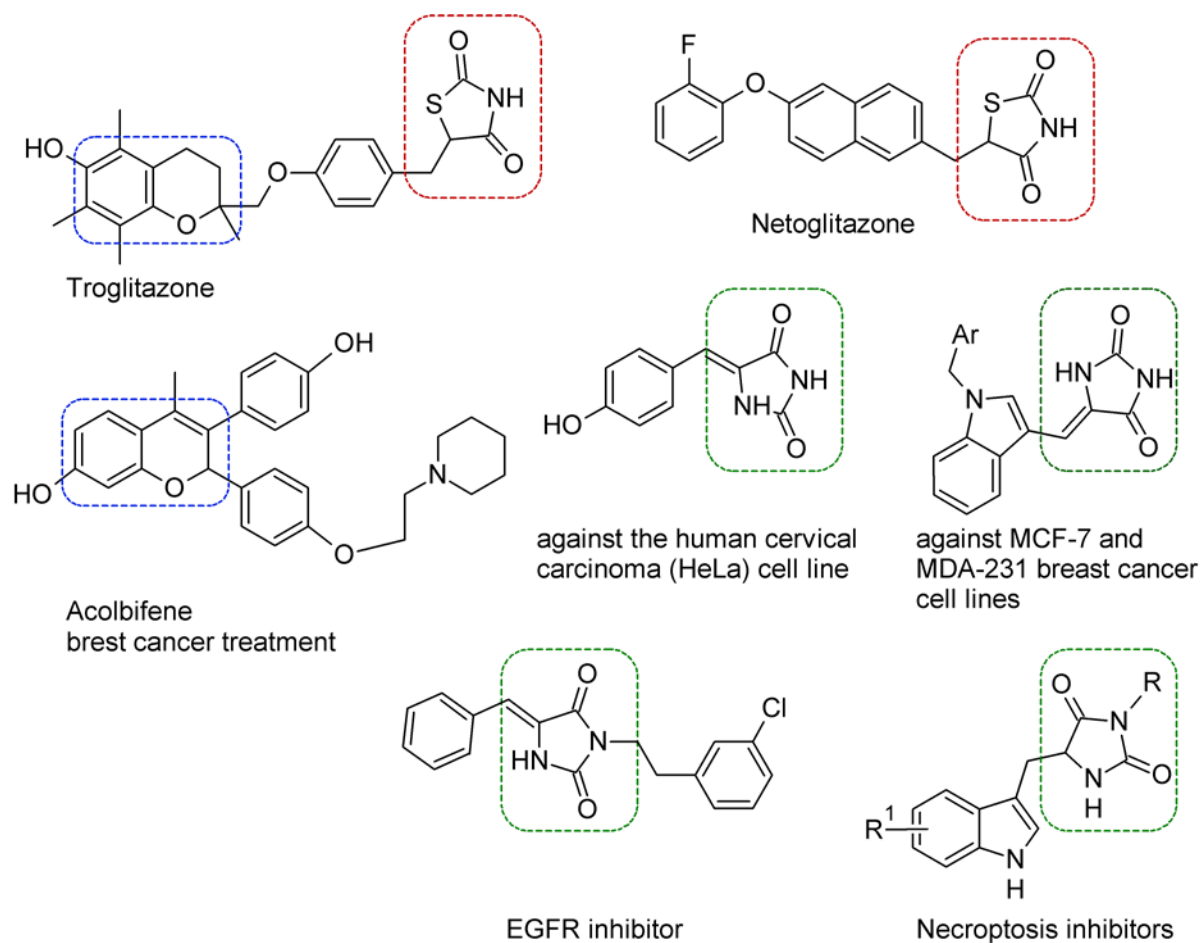
remarkable growth inhibitory activity towards *M. Tuberculosis*. Also, 2-substituted 2*H*-chromenes have wide range of applications [19] as antimycobacterial [20] and antitumor agents [21], inclusive. Therefore, our hypothesis is that chromenyl thiazolidines and hydantoin derivatives might have a broad spectrum of pharmacological activities as well as selective cytotoxicity against cancer cell lines. Continuing the research on the biological activity of 2*H*-chromene derivatives, now we present the synthesis of compounds that comprise a 2*H*-chromene scaffold and thiazolidine-2,4-dione or imidazolidine-2,4-dione heterocyclic rings. Herein we also report on the *in vitro* screening against *M. Tuberculosis* H37Rv as well as antiproliferative/cytotoxic effects of the newly synthesized compounds in a panel of human tumor cell lines.

## EXPERIMENTAL

### General

The IR spectra were recorded on a Nicolet iS10 FT-IR Spectrometer from Thermo Scientific

(USA) using an ATR technique. The NMR experiments on a Bruker Avance II+ 600 MHz NMR spectrometer in DMSO-*d*<sub>6</sub> allowed the assignment of the structures. The precise assignment of the <sup>1</sup>H and <sup>13</sup>C NMR spectra (resolved signals) was accomplished by measurements of 2D homonuclear correlation (COSY), DEPT-135 and 2D inverse detected heteronuclear (C–H) correlations (HMQC and HMBC) and NOESY. HR-ESI-MS spectra were recorded on an LTQ Orbitrap Discovery® spectrometer (ThermoFisher, Germany) equipped with electrospray ionization module Ion Max® (ThermoScientific, USA) operating in positive mode. The melting points were determined using a Buchi 535 apparatus. For TLC was used silica gel 60 GF254 Merck pre-coated aluminum sheets, eluted by hexane-chloroform-acetone 5:3:2 (vol. parts). All reagents were purchased from E. Merck (Darmstadt, Germany) and Aldrich (Milwaukee, MI, USA).



**Fig. 1.** Representative examples of 2*H*-chromene, thiazolidinedione and hydantoin derivatives with anticancer activity and rationale for the designed target compounds.

## SYNTHESIS

**2-Methyl-2H-chromene-3-carbaldehyde 3a** was obtained according to the literature [20]. Yield 0.70 g (57 %), yellow oil. HRMS (ESI) *m/z*: Calcd. [M+H]<sup>+</sup> 175.0753; Found [M+H]<sup>+</sup> 175.07527.

**2-Phenyl-2H-chromene-3-carbaldehyde 3b** was synthesized according to the literature [23] with modifications. In a typical example, 3.19 ml (3.3 g, 25 mmol) cinnamaldehyde, 3.05 ml (3.05 g, 25 mmol) salicylaldehyde, benzoic acid (0.61 g, 5 mmol) and pyrrolidine (0.41 ml, 5 mmol) were dissolved in 100 ml toluene. The mixture was stirred vigorously at 25 °C for 20 hours. The reaction was monitored by thin layer chromatography. The solution was filtered and toluene evaporated on a rotary evaporator at elevated temperature. The residue was dissolved in 5 ml of CHCl<sub>3</sub> and 5 ml hexane, and filtered over 15 g silicagel with eluent hexane–EtOAc mixture 10:1. After eluent evaporation, yellow crystals of the final compound **3b** were obtained. Yield 60%, 3.36 g; yellow crystals; m.p. 75–77 °C; lit. m.p. (23) 75–76 °C. <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>, ppm) δ: 6.257 (s, 1H, H-2), 6.86 (d, *J* = 8.2 Hz, 1H, H-9), 6.996 (dt, *J* = 1.1, 7.5 Hz, 1H, H-7), 7.28–7.31 (m, 5H, Ph), 7.324 (ddd, *J* = 1.6, 7.4, 8.2 Hz, 1H, H-8), 7.457 (dd, *J* = 1.6, 7.5 Hz, 1H, H-6), 7.879 (s, 1H, H-4), 9.673 (s, 1H, CHO). <sup>13</sup>C NMR (150 MHz, DMSO-*d*<sub>6</sub>, ppm): δ 73.63 (C-2), 117.21 (C-9), 120.65 (C-5), 122.44 (C-7), 127.30 (o-Ph), 129.05 (m-Ph), 129.15 (p-Ph), 130.27 (C-6), 133.51 (C-3), 134.12 (C-8), 139.12 (i-Ph), 141.78 (C-4), 154.30 (C-10), 191.56 (CHO). HRMS(ESI) *m/z* found: 237.09076 [M+H]<sup>+</sup>; calcd. for C<sub>16</sub>H<sub>13</sub>O<sub>2</sub>: 237.091006 [M+H]<sup>+</sup>.

**(Z,s-cis)-5-[(2-Methyl-2H-chromen-3-yl)methylidene]-1,3-thiazolidine-2,4-dione 5**. 1,3-Thiazolidine-2,4-dione **4** (3 mmol, 0.35 g) and 2-methyl-2H-chromene-3-carbaldehyde **3** (3 mmol, 0.52 g) were dissolved in abs. ethanol (15 ml). The solution was refluxed for 8 h in the presence of a small amount (0.2 ml) of piperidine as a catalyst. After completion of the reaction (monitored by TLC), the mixture was cooled, the precipitate was filtered and crystallized from ethanol to give the corresponding product as orange-red solid. Yield 0.330 g, (44 %); m.p. 214–215 °C. FTIR cm<sup>-1</sup>: 3350, 1734, 1673, 1607, 1573. <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>): δ (ppm) = 1.304 (d, *J* = 6.5 Hz, 3H, CH<sub>3</sub>), 5.257 (q, *J* = 6.5 Hz, 1H, H-2), 6.858 (d, *J* = 8.0 Hz, 1H, H-5), 6.948 (dt, *J* = 1.0, 7.5 Hz, 1H, H-7), 7.037 (s, 1H, H-4), 7.23–7.26 (m, 1H, H-8), 7.270 (dd, *J* = 1.4, 7.6 Hz, 1H, H-6), 7.291 (s, 1H,

CH H-vinyl), 12.599 (s, 1H, NH). <sup>13</sup>C NMR (150 MHz, DMSO-*d*<sub>6</sub>): δ (ppm) = 19.93 (CH<sub>3</sub>), 71.30 (C-2), 116.45 (C-9), 121.32 (C-S), 121.85 (C-7), 122.61 (C-5), 128.37 (C-8), 128.77 (CH), 130.41 (C-4), 131.59 (C-8), 131.93 (C-3), 151.68 (C-10), 167.22 (C=O), 167.49 (C=O). HRMS (ESI) *m/z*: Calcd. [M+H]<sup>+</sup> 274.05324; Found [M+H]<sup>+</sup>: 274.05334.

**5-[Hydroxy(2-phenyl-2H-1-benzopyran-3-yl)methyl]imidazolidine-2,4-dione 7**. 2-Phenyl-2H-chromene-3-carbaldehyde **3b** (3 mmol, 0.71 g) and imidazolidine-2,4-dione **6** (3 mmol, 0.30 g) were dissolved in abs. ethanol (15 ml). The solution was refluxed for 48 h in the presence of a small amount of piperidine (0.2 ml) as a catalyst. The precipitate thus obtained was collected by filtration and washed with ethanol. Crystallization from methanol and ethyl acetate (1:1) afforded a yellow crystalline product. Yield 0.551 g (54 %); yellow solid; m.p.: 250–251 °C. FTIR cm<sup>-1</sup>: 3227, 1731, 1653, 1603. <sup>1</sup>H NMR (600 MHz, DMSO-*d*<sub>6</sub>): δ (ppm) = 4.200 (dd, *J* = 1.0, 2.3 Hz, 1H, CH-NH), 4.490 (d, *J* = 5.2 Hz, 1H, CHOH), 5.670 (d, *J* = 5.2 Hz, 1H, OH), 6.006 (s, 1H, H-2), 6.630–6.650 (m, 2H, H-9 + H-4), 6.800–6.830 (m, 1H, H-7), 7.020 (dt, *J* = 1.4, 7.7 Hz, 1H, H-8), 7.113 (dd, *J* = 1.4, 7.5 Hz, 1H, H-6), 7.220–7.280 (m, 3H, m-Ph + p-Ph), 7.372 (d, *J* = 6.8 Hz, 1H, o-Ph), 8.035 (s, 1H, CHNH), 10.539 (s, 1H, CONHCO). <sup>13</sup>C NMR (150 MHz, DMSO-*d*<sub>6</sub>): δ (ppm) = 64.21 (CHNH), 71.67 (CHOH), 75.75 (C-2), 116.52 (C-9), 121.22 (C-4), 121.53 (C-7), 122.73 (C-5), 127.18 (C-6), 128.36 (o-Ph), 128.63(m-Ph + p-Ph), 129.60 (C-8), 134.45 (C-3), 139.58 (i-Ph), 151.95 (C-10), 158.10 (NHCONH), 173.61 (CONH). HRMS (ESI) *m/z*: Calcd. [M+H]<sup>+</sup> 337.118283; Found [M+H]<sup>+</sup> 337.11804.

## PHARMACOLOGY

**Antiproliferative activity.** The study was conducted using a panel of cell lines, namely HL-60 (acute myeloid leukemia, established from the peripheral blood of a patient with acute promyelocyte leukemia), K-562 (chronic myeloid leukemia), REH (lymphoid leukemia) and EJ (human urinary bladder carcinoma). EJ cells (also designated MGH-U1) were originally isolated from a high grade (G3) invasive bladder carcinoma. These cell lines had been well validated in our laboratory as a proper test system for platinum agents. The EJ cell line was obtained from the unit of Toxicology and Chemotherapy at the Deutsches Krebsforschungszentrum. The other cell lines were obtained from DSMZ German Collection of Microorganisms and Cell Cultures.

The cell culture flasks and the 96-well microplates were obtained from NUNCLON (Denmark). The stock solutions of the tested compounds (10 mM) were freshly prepared in DMSO. The serial dilutions of the tested compounds were prepared immediately before use. After the final dilutions, the obtained concentrations of DMSO never exceeded 1%. Cytotoxicity of the compounds was assessed using the MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] dye reduction assay as described by Mossman [24] with some modifications [25]. Exponentially growing cells were seeded in 96-well microplates (100 IL/well at a density of 3.5/10<sup>5</sup> cells/mL for the adherent and 1/10<sup>5</sup> cells/mL for the suspension cell lines) and allowed to grow for 24 h prior the exposure to the studied compounds. The cells were exposed to the tested agents for 72 h, whereby a set of 8 separate wells was used for each concentration. Every test was run in triplicate. After incubation with the tested compounds MTT solution (10 mg/mL in PBS) aliquots were added to each well. The plates were further incubated for 4 h at 37 °C and the formazan crystals formed were dissolved by adding 110 IL of 5% HCOOH in 2-propanol. The MTT-formazan absorption was measured using a multimode microplate reader (Beckman Coulter DTX880) and the results were normalized as a percentage of the untreated control (set as 100% viable). The MTT-bioassay data were normalized as a percentage of the untreated control (set as 100 % viability), were fitted to sigmoidal dose response curves and the corresponding IC<sub>50</sub> values (concentrations causing 50 % suppression of cellular viability) were calculated using non-linear regression analysis (Curve-fir; GraphPad Prism software for PC).

The cell lines were purchased from the DSMZ GmbH, (Braunschweig, Germany). They were cultured under standard conditions - RPMI-1640 medium supplemented with 10% fetal bovine serum (FBS) and 2 mM L-glutamine - in cell culture flasks housed at 37 °C in an incubator, BB16-Function Line<sup>®</sup> Heraeus (Kendro, Germany) with humidified atmosphere and 5 % of CO<sub>2</sub>. The cell cultures were maintained in log phase by supplementation with fresh medium two or three times weekly.

*Antimycobacterial activity.* The antimycobacterial activity was determined towards reference strain *M. Tuberculosis* H37Rv through the proportional method of Canetti [26]. This method, recommended by WHO, is the one most used worldwide for exploration of

sensitivity/resistance of tuberculosis strains towards chemotherapeutics. It allows precise determination of the proportion of mutants resistant to a certain drug. A sterile suspension/solution of the tested compound was added to Löwenstein-Jensen based medium before its coagulation (30 min at 85 °C). The compound was tested at four concentrations – 2 µg/ml, 0.2 µg/ml, 0.1 µg/ml and 0.05 µg/ml, (in DMSO). Tubes with Löwenstein-Jensen medium (5 ml) containing the tested compounds and those without them (controls) were inoculated with a suspension of *M. tuberculosis* H37Rv (10<sup>5</sup> cells/ml) and incubated for 45 days at 37 °C. The ratio between the number of colonies of *M. tuberculosis* grown in medium containing the compounds and the number of colonies in the control medium were calculated and expressed as a percentage of inhibition. The cell culture flasks and the 96-well microplates were obtained from NUNCLON (Denmark). The stock solutions of the tested compounds (10 mM) were freshly prepared in DMSO. The serial dilutions of the tested compounds were prepared immediately before use. After the final dilutions the obtained concentrations of DMSO never exceeded 1%. Cytotoxicity of the compounds was assessed using the MTT [3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide] dye reduction assay as described by Mossman [24] with some modifications [25]. Exponentially growing cells were seeded in 96-well microplates (100 IL/well at a density of 3.5/10<sup>5</sup> cells/ mL for the adherent and 1/10<sup>5</sup> cells/mL for the suspension cell lines) and allowed to grow for 24 h prior the exposure to the studied compounds. Cells were exposed to the tested agents for 72 h, whereby a set of 8 separate wells was used for each concentration. Every test was run in triplicate. After incubation with the tested compounds MTT solution (10 mg/mL in PBS) aliquots were added to each well. The plates were further incubated for 4 h at 37 °C and the formazan crystals formed were dissolved by adding 110 µl solvent (5% HCOOH in 2-propanol). The MTT-formazan absorption was measured using a multimode microplate reader (Beckman Coulter DTX880) and the results were normalized as a percentage of the untreated control (set as 100% viable). The MTT-bioassay data were normalized as a percentage of the untreated control (set as 100 % viability), were fitted to sigmoidal dose response curves and the corresponding IC<sub>50</sub> values (concentrations causing 50 % suppression of cellular viability) were calculated using non-linear regression analysis (Curve-fir; GraphPad Prism software for PC).



The MIC was defined as the minimum concentration of the compound required to inhibit bacterial growth completely (0% growth). The MIC values were calculated and given as  $\mu\text{M}$ . Ethambutol and isoniazid were used as controls.

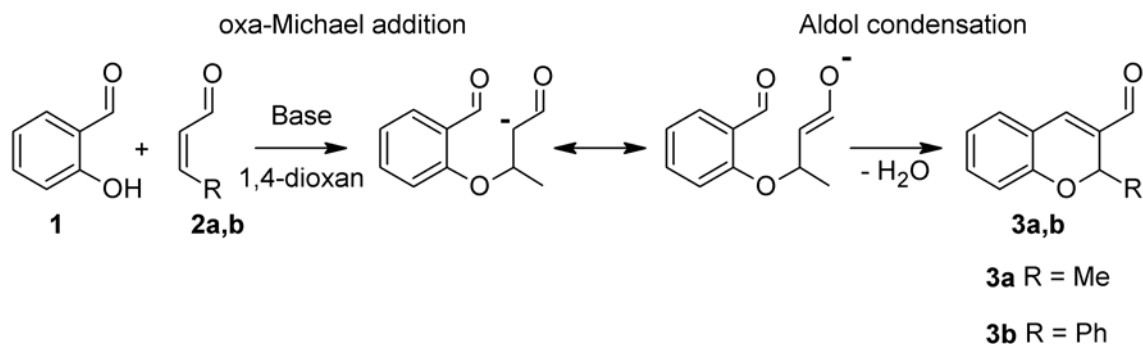
## RESULTS AND DISCUSSION

**Synthesis.** The synthetic route for preparing the compounds was illustrated in Schemes 1 and 2. 2*H*-Chromene derivatives **3a,b** were selected as starting compounds and were synthesized *via* Michael-aldol reaction, according to the known procedure [27] (Scheme 1). The reaction of 2-hydroxybenzaldehyde with  $\alpha,\beta$ -unsaturated aldehydes under basic conditions afforded appropriate 2-substituted 2*H*-chromene-3-carbaldehyde **3a,b**. The preparation of 2-methyl-2*H*-chromene-3-carbaldehyde **3a** is conducted in 1,4-dioxane under reflux. For the synthesis 2-

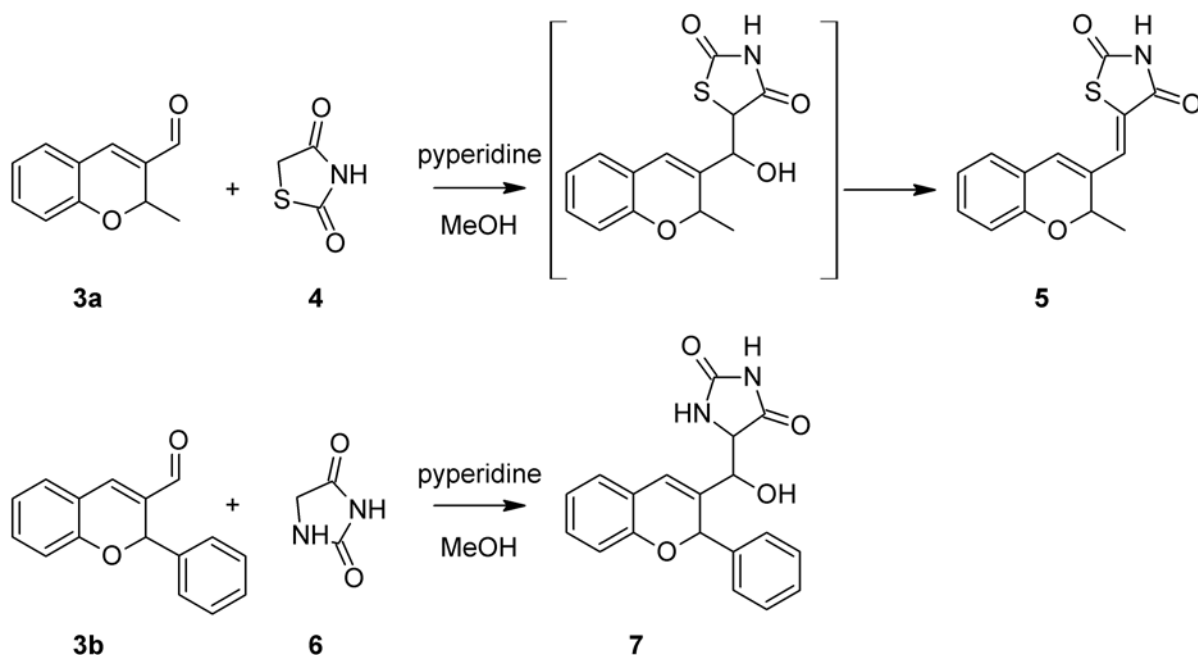
phenyl-2*H*-chromene-3-carbaldehyde **5** we used piperidine as a catalyst [23].

The new compound 5-[(2-methyl-2*H*-chromen-3-yl)methylidene]-1,3-thiazolidine-2,4-dione **5** was synthesized by Knoevenagel condensation of 2-methyl-2*H*-chromene-3-carbaldehyde **2a** and 1,3-thiazolidine-2,4-dione **3** (Scheme 2).

Several conditions were tested (sodium acetate in acetic acid, sodium acetate in DMF and piperidine in ethanol/methanol under thermal conditions) for the preparation of 2-methylchromene - derivative **5**, but the best result (a moderate yield of 44 %) was obtained by refluxing ethanol and in the presence of a catalytic amount of piperidine. As outlined in Scheme 2, when 2-phenyl-2*H*-chromene-3-carbaldehyde **5** reacted with hydantoin **6** under the selected



**Scheme 1.** Synthesis of 2-methyl-2*H*-chromene-3-carbaldehyde **3a** and 2-phenyl-2*H*-chromene-3-carbaldehyde **3b** through the domino oxa-Michael/aldol condensation reactions.



**Scheme 2.** Synthesis of 2-methylchromene derivative **5** and 2-phenylchromene derivative **7**.

conditions, dehydration did not occur and the final product is intermediate alcohol 5-[hydroxy(2-phenyl-2*H*-chromen-3-yl)methyl]imidazolidine-2,4-dione **7**. Our efforts to obtain a Knoevenagel product after the condensation reaction under the conditions described above failed and suggested that other factors might influence the reaction.

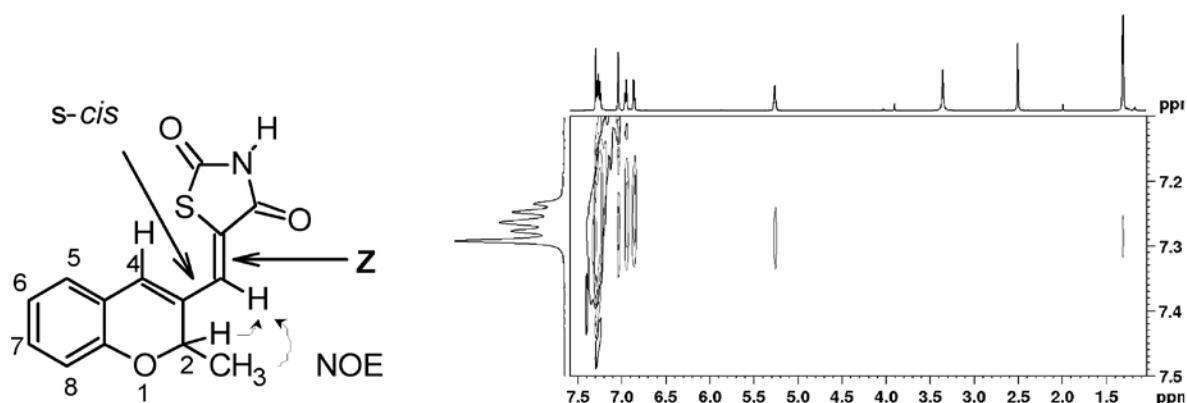
The structures of novel compounds **5** and **7** were proven by means of FTIR, <sup>1</sup>H, <sup>13</sup>C NMR and HR-ESI-MS spectral methods and were confirmed additionally by extensive two-dimensional (2D) NMR (COSY, multiplicity-edited HSQC, HMBC) and NOESY spectra. In the <sup>1</sup>H NMR in DMSO-*d*<sub>6</sub> spectrum of product **5** the signal for CH<sub>2</sub> protons in thiazolidinedione ring is absent and in addition to the aromatic protons of chromene ring (6.858-7.291 ppm), a sharp singlet due to vinyl proton is observed at 7.291 ppm. The <sup>13</sup>C NMR spectrum of 2-methylchromene derivative **4** shows 13 signals. The signal for C-2 of chromene ring appears at 71.30 ppm, the vinyl proton resonates at 151.68 ppm and the signals for the carbonyl are observed at 167.22 and 167.49, respectively. Meanwhile, in the <sup>1</sup>H NMR spectra of 2-methylchromene derivative **5** only a set of signals occurs what confirms that only one stereoisomer *Z* has been obtained during the condensation. That is in agreement with the literature [27] and isomer *Z* appears to be more thermodynamically stable than isomers *E*. The NOESY spectrum (Fig. 2) shows a NOE effect and proximity between the vinyl proton and H-2; and between the vinyl proton and CH<sub>3</sub> protons. That proves the (*s-cis*)-conformation of the two conjugated double bonds. The described 2-methylchromene derivative **5** possessing a chiral center at C-2 position is a racemate.

In the <sup>1</sup>H NMR spectrum of product **7** in DMSO-*d*<sub>6</sub>, there is no signal for CH<sub>2</sub> protons in the hydantoin ring. In addition to the aromatic protons of chromene ring (6.630-7.372 ppm), a doublet of doublets at 4.200 for CH-NH and a broad doublet at 4.490 (*J*=5.2 Hz) for CHOH are observed. A doublet at 5.670 for the hydroxyl group with the same coupling constant of 5.2 Hz is registered. The lack of a cross peak for this signal in HSQC spectrum for OH proton, indicates that there is not a carbon atom connected directly to it. Having localized the position of OH proton it is easy to assign the remaining protons from COSY spectrum. All carbon signals are assigned using HSQC and HMBC. The signal at 4.200 ppm is coupled by two

small *J* constants (1.0 and 2.3 Hz). One of these constant is a *J* constant to the NH proton, the other is a *gauche* constant in the 4-substituted ethane structure. The infrared spectrum of 2-methylchromene derivative **5** shows a strong absorption at 1733 cm<sup>-1</sup> and 1678 cm<sup>-1</sup> corresponding to C=O functional groups, a band at 1607 cm<sup>-1</sup> (C=C stretching in chromene ring) and 1573 cm<sup>-1</sup> corresponding to C=C bond. The IR spectrum of compound **7** shows bands at 3227 cm<sup>-1</sup> (NH stretching), 1731 and 1698 cm<sup>-1</sup> (C=O functional groups) and 1603 cm<sup>-1</sup> (C=C stretching in chromene ring). In addition, the novel structures are supported by a positive HR-ESI-MS spectrum which reveals a molecular ion peak at *m/z* 274.05334 [M+H]<sup>+</sup>, (calcd. [M+H]<sup>+</sup> 274.05324) and 337.11804 [M+H]<sup>+</sup> (calcd. [M+H]<sup>+</sup> 337.118283) and allows confirming the molecular formulas, C<sub>14</sub>H<sub>11</sub>NO<sub>3</sub>S and C<sub>19</sub>H<sub>16</sub>N<sub>2</sub>O<sub>4</sub>, of compounds **5** and **7**, respectively.

*Antiproliferative/cytotoxic effects.* Table 1 summarizes the antiproliferative effects of 5-[(2-Methyl-2*H*-chromen-3-yl)methylidene]-1,3-thiazolidine-2,4-dione **5** and 5-[Hydroxy(2-phenyl-2*H*-1-benzopyran-3-yl)methyl]imidazolidine-2,4-dione **7** against a panel of four human tumor cell lines, namely HL-60 (acute promyelocyte leukemia), REH (lymphoid leukemia), K-562 (chronic myeloid leukemia) and EJ (urinary bladder carcinoma), using the alkylating agent melphalan as a reference anticancer drug.

The results of the *in vitro* cytotoxicity bioassay indicated that the compounds **5** and **7** exerted moderate to potent growth inhibition against the tested cancer cells with IC<sub>50</sub> values of 13.1- 98 μM. Compound **5** showed higher activity than compound **7** against HL-60, REH and EJ with the IC<sub>50</sub> values of 39.6, 13.1 and 26.2 μM, respectively. As for the activity against chronic myeloid leukemia K-562, the highest cytotoxic activity was displayed by compound **7** with IC<sub>50</sub> a value of 83.7 μM. Additionally, compound **5** was found to be more potent against REH tumor cell lines (IC<sub>50</sub> = 13.1 μM) than Melphalan. REH cells were the most sensitive to the studied compounds and the potency of the most active compounds is similar to that of Melphalan. EJ cells seem to be the most resistant to compound **7**. However, compound **5** showed a relatively high antiproliferative potency toward EJ cells.



**Fig. 2.** Part of NOESY spectrum of 2-methylchromene derivative **5** showing the NOE effects between vinyl proton at 7.291 and 5.257 (H-2) ppm, and between 7.291 and 1.304 (CH<sub>3</sub>) ppm, which confirms the (*s-cis*)-conformation.

**Table 1.** Cytotoxicity (IC<sub>50</sub>) of the compounds **5** and **7** against different cell lines and antimycobacterial activity against reference strain *Mycobacterium tuberculosis* H37Rv.

Compd	Structure	IC <sub>50</sub> (μM) <sup>1</sup>				MIC <sup>6</sup> (μM)
		HL-60 <sup>2</sup>	K-562 <sup>3</sup>	REH <sup>4</sup>	EJ <sup>5</sup>	
5		39.6 ± 2.7	98.0 ± 3.9	13.1 ± 2.4	26.2 ± 2.5	0.360
7		73.2 ± 4.8	83.7 ± 3.6	26.3 ± 2.8	117.5 ± 4.0	0.297
Melphalan <sup>7</sup>		11.2 ± 1.9	28.3 ± 3.2	14.6 ± 3.1	13.7 ± 1.9	-
EMB_2HCl <sup>8</sup>		-	-	-	-	1.45
INH <sup>9</sup>		-	-	-	-	0.64

<sup>1</sup>Means±sd from eight independent experiments. Cell line: <sup>2</sup>HL-60 (acute promyelocyte leukemia), <sup>3</sup>K-562 (chronic myeloid leukemia), <sup>4</sup>REH (lymphoid leukemia) and <sup>5</sup>EJ (urinary bladder carcinoma); <sup>6</sup>reference strain of *Mycobacterium tuberculosis* H37Rv, MIC (μM) was defined as the lowest concentration resulting in a complete inhibition of the bacterial growth and reproduction; <sup>7</sup>Melphalan - reference compound; <sup>8</sup>Isoniazid - reference compound and <sup>9</sup>EMB-2HCl (ethambutol dihydrochloride) - reference compound.

**Antimycobacterial activity.** The *in vitro* antimycobacterial activity of all compounds against *Mycobacterium tuberculosis* H37Rv reference strain was evaluated using the proportional method of Canetti and the resazurin microtiter assay. Ethambutol - EMB-2HCl (ethambutol dihydrochloride) and isoniazid were used as controls. The MIC is defined as the lowest concentration effecting the 100 % reduction in fluorescence, relative to controls. The data for the compounds **5** and **7** are shown in Table 1. The tested compounds exhibited significant antimycobacterial activity against the chosen strain,

whereby the MIC values were within the nanomolar range, lower than those of ethambutol and INH.

## CONCLUSION

New thiazolidine-2,4-dione and hydantoin derivatives were synthesized and their structures were elucidated on the basis of FTIR, <sup>1</sup>H NMR, <sup>13</sup>C NMR, 2D spectra (COSY, HMQC, HMBC) and HRMS data. The aldol product **5** was exclusively obtained in (*Z*)-configuration. The compounds **5** and **7**, were tested for cytotoxic activity with MTT-dye reduction assay against leukemia-derived HL-60 cells, REH, K-562, and urinary bladder

carcinoma cells (EJ). The tested compounds displayed promising micromolar antiproliferative activity. The most potent compound was 2*H*-chromene derivative **5** with IC<sub>50</sub> values ranging from 13.1 - 0.98 μM. Also, the tested compounds **5** and **7** were found highly potent against the *M. tuberculosis* H37Rv, demonstrating a nanomolar activity (MICs ranging from 0.27 to 0.71 μM). The obtained results could be useful for the design and synthesis of new substituted 2,4-thiazolidindione with superior antiproliferative potency as potential anticancer agents. Taken together our data give the reason to consider 2,4-imidazolinone derivative **5** and hydantoin derivative **7** promising new leads for further exploration as potential antimycobacterial agents.

#### REFERENCES

1. A. Smarandache, V. Nastasa, M. Boni, A. Staicu, J. Handzlik, K. Kiec-Kononowicz, L. Amaral, M.-L. Pascu, *Physicochemical and Engineering Aspects*, **505**, 37 (2016).
2. A.A. Napoleon, *International Journal of PharmTech Research*, **9**(3), 429 (2016).
3. V.S. Jain, D.K. Vora, C.Ramaa, *Bioorganic & medicinal chemistry*, **21**(7), 1599 (2013).
4. H.L.T. Anh, N.T. Cuc, B.H. Tai, P.H. Yen, N.X. Nhiem, D.T. Thao, N.H.Nam, C.V. Minh, P.V. Kiem, Y.H. Kim, *Molecules*, **20**(1):1151 (2015).
5. R. Romagnoli, P.G. Baraldi, M.K. Salvador, M.E. Camacho, J. Balzarini, J. Bermejo, J. bermejo, F. Estevez, *European journal of medicinal chemistry*, **63**, 544 (2013).
6. K.A. Szychowski, M.L. Leja, D.V. Kaminsky, U.E. Binduga, R. Pinyazhko, R.B. Lesyk, J. Gminski, *Chemico-Biological Interactions*, **262**, 46 (2017).
7. H. Joshi, T. Pal, C. Ramaa, *Expert opinion on investigational drugs*, **23**(4), 501 (2014).
8. C. Carmi, A. Cavazzoni, V. Zuliani, A. Lodola, F. Bordi, P.V. Plazzi, R.R. Alfieri, P.G. Petronini, M. Mor, *Bioorganic & medicinal chemistry letters*, **16**(15), 4021 (2006).
9. Z. Rajic, B. Zorc, S. Raic-Malic, K. Ester, M. Kralj, K. Pavelic, et al., *Molecules*, **11**(11), 837 (2006).
10. L. Sacconnay, L. Ryckewaert, G.M. Randazzo, C. Petit, C.D.S. Passos, J. Jachno, V. Michailoviene, A. Zubriene, D. Matulis, P.-A. Carrupt, C.A. Simoes-Pires, A. Nurisso, *European Journal of Pharmaceutical Sciences*, **85**, 59 (2016).
11. M. Mudit, M. Khanfar, A. Muralidharan, S. Thomas, G.V. Shah, R.W. van Soest, K.A. El Sayed, *Bioorganic & medicinal chemistry*, **17**(4), 1731 (2009).
12. Y.T. Reddy, P.N. Reddy, S. Koduru, C. Damodaran, P.A. Crooks, *Bioorganic & medicinal chemistry*, **18**(10), 3570 (2010).
13. X. Teng, A. Degtarev, P. Jagtap, X. Xing, S. Choi, R. Denu, J. Yuan, G.D. Cuny, *Bioorganic & medicinal chemistry letters*, **15**(22), 5039 (2005).
14. D.T. Youssef, L.A. Shaala, K.Z. Alshali, *Marine drugs*, **13**(11), 6609 (2015).
15. K. Ozadali, O.U. Tan, P. Yogeewari, S. Dharmarajan, A. Balkan, *Bioorganic & medicinal chemistry letters*, **24**(7), 1695 (2014).
16. F.M. Shaikh, N.B. Patel, G. Sanna, B. Busonera, P. La Colla, D.P. Rajani, *Medicinal Chemistry Research*, **24**(8), 3129 (2015).
17. D. Łażewska, P. Maludziński, E. Szymańska, K. Kieć-Kononowicz, *Biomedical Chromatography*, **21**(3), 291 (2007).
18. Y. Liu, W. Zhong, S. Li, *Chinese Chemical Letters*, **23**(2), 133 (2012).
19. N. Thomas, S.M. Zachariah, *Asian J Pharm Clin Res.*, **6**(2), 11 (2013).
20. V.T. Angelova, V. Valcheva, N.G. Vassilev, R. Buyukliev, G. Momekov, I. Dimitrov, et al., *Bioorganic & Medicinal Chemistry Letters*, **27**(2), 223 (2017).
21. S. Rahmani-Nezhad, M. Safavi, M. Pordeli, S.K. Ardestani, L. Khosravani, Y. Pourshojaei, et al., *European journal of medicinal chemistry*, **86**, 562 (2014).
22. V.T. Angelova, N.G. Vassilev, B. Nikolova-Mladenova, J. Vitas, R. Malbaša, G. Momekov, L. Saso, *Medicinal Chemistry Research*, **25**(9), 2082 (2016).
23. I. Ibrahim, H. Sundén, R. Rios, G.-L. Zhao, A. Córdoba, *CHIMIA International Journal for Chemistry*, **61**(5), 219 (2007).
24. T. Mosmann, *Journal of immunological methods*, **65**(1-2), 55 (1983).
25. S.M. Konstantinov, H. Eibl, M.R. Berger, *British Journal of Haematology*, **107**(2), 365 (1999).
26. G. Canetti, W. Fox, A. Khomenko, H. Mahler, N. Menon, D. Mitchison, et al., *Bulletin of the World Health Organization*, **41**(1), 21 (1969).
27. M. Azizmohammadi, M. Khoobi, A. Ramazani, S. Emami, A. Zarrin, O. Firuzi, et al., *European journal of medicinal chemistry*, **59**, 15 (2013).



## СИНТЕЗ, *in vitro* АНТИПРОЛИФЕРАТИВНА И АНТИМИКОБАКТЕРИАЛНА АКТИВНОСТ НА ТИАЗОЛИДИН-2,4-ДИОН И ХИДАНТОИНОВИ ПРОИЗВОДНИ

В. Т. Ангелова<sup>1,\*</sup>, В. Вълчева<sup>2</sup>, Н. Василев<sup>3</sup>, Р. Буюклиев<sup>1</sup>, Р. Михайлова<sup>1</sup>, Г. Момеков<sup>1</sup>

<sup>1</sup>Фармацевтичен факултет, Медицински университет София, ул. Дунав 2, 1000 София, България;

<sup>2</sup>Институт по микробиология "Стефан Ангелов", Българска академия на науките, ул. Акад. Бончев 26, 1113 София, България;

<sup>3</sup>Институт по органична химия с Център по фитохимия, Българска академия на науките, ул. Акад. Бончев 9, 1113 София, България

Получена на 23 февруари 2017 г.; приета на 27 април 2017 г.

(Резюме)

Синтезирани са нови 2*H*-хроменови производни, съдържащи тиазолидин-2,4-дионов и хидантоинов фрагменти и структурата им е потвърдена чрез <sup>1</sup>H NMR, <sup>13</sup>C NMR, 2D NMR, FTIR спектрални методи и HR-ESI-MS. Съединенията са изследвани *in vitro* за цитотоксичната им активност срещу четири човешки туморни клетъчни линии - HL-60 (остра промиелоцитна левкемия), REN (лимфоидна левкемия), K-562 (хронична миелоидна левкемия) и EJ (карцином на пикочния мехур). 2*H*-Хромен съдържащият тиазолидин-2,4-дион **5** е много активен срещу три ракови клетъчни линии, с IC<sub>50</sub> от порядъка на 13.1-39.6 μM и показва най-изразена антипролиферативна активност срещу лимфоидна левкемия (клетъчна линия REN) със стойност на IC<sub>50</sub> = 13.1 μM. Хидантоин съдържащото 2*H*-хроменово производно **7** е високо най-ефективно срещу хронична миелоидна левкемия K-562 със стойност на IC<sub>50</sub> = 83.7 μM. Съединенията **5** и **7** демонстрират също и значителна antimycobacterial активност срещу *Mycobacterium tuberculosis* H37Rv с минимална инхибираща концентрация (MIC) в диапазона от 0.29 и 0.36 μM, съответно.