A new study on the derivatization of malondialdehyde by anisidine reagent with use of UV-Vis, FT-IR, and NMR techniques

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Received June 26, 2015, Revised September 10, 2015

Derivative methods of malondialdehyde (MDA) as a biomarker of oxidative stress often require high temperature and long heating time. Some of which activate a color reaction such as thiobarbituric acid reactive substances (TBA), Diaminonaphtalene, Pentafluorophenyl hydrazine, and Phenylhydrazine after placing in that harsh condition. Our attempts were to introduce a new derivative reaction between malondialdehyde and Para methoxy aniline (PMA) to identify MDA. The derivatization of malondialdehyde (MDA) with different concentration of para-methoxy aniline (Anisidine) in an acidic medium and various time, temperature, and pH was investigated. The possible reaction of malondialdehyde and Anisidine was completed after 10 min at 75oC in optimal condition. By spectrophotometric techniques, the complex formed in this process exhibits a highly specific UV spectrum with a sharp maximum at 400nm. (1H and 13C) NMR, and FT-IR were confirmed the formed adduct.

**Keywords:** Malondialdehyde, Anisidine, colorimetric reaction, derivatization

1. **INTRODUCTION**

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Malondialdehyde (MDA), as one of the great significance individual aldehyde resulting from lipid peroxidation, is a reactive unsaturated dicarbonyl that can easily bind to macromolecules such as structural and functional proteins and nucleic acids (Karatas *et al*., 2002; Grotto *et al*., 2007; Del Rio et al., 2005; Agarwal *et al*.,2007; Seljeskog *et al.,* 2006). As of now, the great number of analytical approaches for determination of Malondialdehyde (MDA) with various derivative methods have been proposed. Most procedures based on the aldehydic reactivity of MDA employed hydrazine-based derivatization reagents (Giera *et al.,*2012; Grintzalis *et al.,* 2013). Still, the classical assay "2-thiobarbituric acid (TBA) is the most general applied technology in which two molecules of TBA change with one molecule of MDA to give a colored reaction product which can be measured spectrophotometrically at 535 nm, or by fluorescence detection with excitation at 530 nm and emission at 550 nm(Steghens *et al.,* 2001; Czauderna *et al.,* 2011; Mendes *et al.,* 2009; Pilz *et al.,* 2000; Shibamoto *et al.,* 2006). In fact, the TBA assay is a non-specific method for MDA; therefore, further developed methods were strongly suggested by researchers with the use of new color reagent for detection of this prominent biomarker. Sim *et al.,* 2003; Khoubnasabjafari *et al.,* 2015; Shahidi, F. *et al.,* 2005). Derivative reagents, namely, thiobarbituricacid, diamino-naphtalene (DNPH), Pentafluorophenylhydrazine (PFH), Phenylhydrazine (PH), dansylhydrazine, methylhydrazine, 2,2,2-Trifluoroethylhydrazine and FMOC-hydrazine after placing in high temperature, and longtime have been proposed by researchers to measure MDA through expensive separation technology such as reverse phase**-**liquid chromatography (RP-HPLC), gas chromatography–mass spectrometry (GC-MS), and liquid chromatography–mass spectrometry (LC-MS, or alternatively HPLC-MS) (Giera *et al.,*2012). However*,* the resultof their investigations was unsatisfactory***.***

On the other hand, *para*-anisidine (p-anisidine) value would be a well-known method to measure the content of aldehydes (principally 2 alkenals and 2, 4-alkadienals) generated during the decomposition of hyroperoxides. *para*-methoxyaniline (anisidine or PMA) and the aldehydic compounds under acidic conditions provide yellowish products and absorb at 350 nm (Aruoma *et al.,*1998). Besides, p-An Value is a valid indicator of oxidative rancidity in oils and fatty foods (Mao *et al.,* 2006; Shibamoto *et al.,*2006). However, no specific method has been developed for determination of malondialdehyde by anisidine reagent.

In this paper, based on colorimetric reaction, a new method for synthesis and identification of MDA was introduced. Investigation of complex formation (MDA-PMA) with the use of variegated techniques was studied as well. In addition, it was expected that this procedure would assure a simple, selective, and sensitive measurement of MDA for further biological studies.

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**2. EXPERIMENTAL**

***2.1. Material and methods***

Malondialdehyde Tetra butyl ammonium salt (MDA, 96%) was purchased from Sigma Aldrich. In addition, para methoxy aniline, butylatedhydroxytoluene, glacial acid acetic, HClO4 0.1 M diluted in glacial acetic acid, absolute ethanol, and sodium hydroxides were from Merck Company.

***2.2. Solvent selection***

To find an appropriate solvent, a possible reaction between Malondialdehyde Tetra butyl ammonium salt (MDA, 96%) and para-methoxy aniline (anisidine, PMA) was evaluated. Therefore, different solvents such as HPLC-grade water, absolute ethanol, glacial acid acetic, and HCl were examined. Finally, absolute ethanol and glacial acetic acid were selected as two proper solvents for MDA and Para methoxy aniline, respectively.

*2.3. Preparation of Anisidine reagent*

*p*-methoxy aniline (Anisidine) as a reagent was prepared at 1000 µM.L-1(µM = µM.L-1 =n mol/ ml) in glacial acetic acid, by dissolving 0.0123 g of ansidine in 100 ml of glacial acetic acid. The solution was protected from the light and was stored at room temperature for a daily basis. The maximum absorbance was at 273 nm (Figure.1).



**Fig. 1.** Wavelength of Anisidine diluted in glacial acetic acid

Anisidine was fully tested on room temperature in the dark place with two different concentrations (500 and 1000 µM) to investigate its stability in long period. Anisidine stability had a constant trend within 30 days.

***2.4. Preparation of MDA stock solution***

Standard stock solution of MDA (5000 µM.L-1) was obtained by dissolving 0.0156 g of Malondialdehyde Tetra butyl ammonium salt in 10ml of absolute ethanol. **(**Figure.2**).** This stock solution remained stable for at least 6 months, and stored at -20 oC in aliquots of 250 µl in dark place while the working standard solutions had to be prepared every day MDA solution was monitored by its absorbance at 267nm (Giera *et al.,*2012).

***2.5. The process of derivatisation***

First, seven clean glass test tubes were taken and labeled them (A-G). Second, different concentrations of MDA standard stock solutions were prepared (50, 25, 10.5, 5, 2.5, 1, and 0.5 μM) and diluted by absolute ethanol. Afterwards, two ml of prepared MDA solution was transferred to each labeled glass tubes. Then, two ml of Anisidine (500 μM) as a color reagent was added to them and vortex the solution. Finally, the tubes were closed and placed them to any holder to keep the tubes upright during boiling at 90 oC. After 10 min, immediately, the tubes were removed and were placed in ice bath to stop the reaction. At least, 15 min was needed to be cooled down. Without any extraction steps, two ml of the clear yellowish complex of each glass tubes were loaded to absorption cell for reading the absorbance at 400-404 nm by UV-Vis.



**Fig. 2.** Wavelength of malondialdhyde tetrabutyl ammonium salt diluted in absolute ethanol.

***2.5. The process of derivatisation***

First, seven clean glass test tubes were taken and labeled them (A-G). Second, different concentrations of MDA standard stock solutions were prepared (50, 25, 10.5, 5, 2.5, 1, and 0.5 μM) and diluted by absolute ethanol. Afterwards, two ml of prepared MDA solution was transferred to each labeled glass tubes. Then, two ml of Anisidine (500 μM) as a color reagent was added to them and vortex the solution. Finally, the tubes were closed and placed them to any holder to keep the tubes upright during boiling at 90 oC. After 10 min, immediately, the tubes were removed and were placed in ice bath to stop the reaction. At least, 15 min was needed to be cooled down. Without any extraction steps, two ml of the clear yellowish complex of each glass tubes were loaded to absorption cell for reading the absorbance at 400-404 nm by UV-Vis.

***2.6. Investigation of concentration, time, and temperature***

Various concentrations of MDA standards (50, 12.5, 3.125 µM.L-1) and Anisidine reagent (30, 50, 100, 150, 250, 500 µM.L-1) were prepared by further dilution of their stock solutions. After preparing anisidine reagent in glacial acetic acid, the solution should be protected from the light within the procedure. For activating the color, derivatization was performed at different reaction times (5, 10, 20, 30, and 40 min) and at different temperature (30, 70, and 90 oC) to evaluate the optimum conditions of MDA derivatization.

***2.7. Process of complex (MDA-PMA) formation***

Appropriate volumes and concentrations of anisidine, optimum temperature, and time were the significant parameters that needed to be studied. In fact, to explore the colorimetric reaction between MDA and PMA, several tests with various concentrations of each have been done. Briefly, by our research work, various concentrations of MDA standards (50, 12.5, 3.125 µM) and anisidine reagent (150, 250, 500 µM) were prepared by further dilution of the stock solutions. Afterwards, two milliliter of prepared MDA solution (50) was transferred to a labeled glass tube (Complex MDA-PMA).Then, two ml of Anisidine (500 μM) as a color reagent was added to the labeled tube (Complex MDA-PMA) and vortex the solution. Finally, the tube was closed and placed them to any holder to keep the tubes upright during boiling at 90, 70, and 30 oC . After 10 min, immediately, the tubes were removed and placed in ice bath to stop reaction. At least, 10 min was needed to be cooled down. The optimum condition provided for stable yellowish color adduct (complex A) was considered as follows: (Time: 20 min, Temperature 75 oC , Anisidine concentration 500 µM).

***2.8. Synthesis of MDA-PMA adduct***

The product (MDA-PMA) is not commercially available, but it can be prepared in laboratory condition. Formation of imine compound was necessary to evaluate the reaction and confirmation of MDA-PMA adduct. The derivatisation procedure is based on the principle of (0.5g) of MDA reacting with (0.4g) of anisidine prepared with glacial acetic acid at room temperature.

It was found that an imine compound is produced after a few seconds. Then, yellow-orange product was dried at 60 oC for 4 h in a vacuum drying oven. The structure of this compound was confirmed by different methods namely NMR (1H and 13C), and FT-IR.

***2.9. Effect of pH***

The pH effect of the derivatization reaction on the MDA value was investigated by previous scientists [3]. Although this new procedure was confirmed at pH= 7.4, the validity of the derivative method has been questioned in various pH (acidic or basic medium). In other words, the complete reaction should be evaluated in different pH for the future biological studies. For this reason, the pH ranges (3, 7, and 12.0) have been selected.

In normal pH (p H= 7.4), two ml of MDA (50 µM) was added to two ml of Anisidine (500µM) in a 10 ml lab tube. Then, this reaction mixture was placed in the water bath for 10 min at 90 oC. The solution was cooled and ready for the analysis by UV-Vis spectrophotometer at 400-404nm.

In the basic medium, 200 µl of 6 M NaOH was added to 1000 µl of MDA in a 10ml lab tube. By placing this mixture at 60 oC in the water bath for 30 min, allow medium to be basic. After cooling, 200 µl of Anisidine reagent (500µM) was added to this basic solution. The same as above, the reaction mixture was placed in water bath for 10 min at 90 oC. The solution was cooled and ready for the analysis by UV-Vis spectrophotometer at 400-404nm.

In the acidic medium, 250 µl of HClO4 (0.1 M) diluted in glacial acetic acid was added to 1000 µl of MDA (50 µM) in a 10ml lab tube and vortex. Then, 200 µl of Anisidine reagent (500µM) was added to the tube and finally, this mixture was placed in water bath for10 min at 90 oC. The solution was cooled and ready for the analysis by spectrophotometer at 400 nm.

***2.10. Techniques***

Possible structure of the formed adduct (complex MDA-PMA) under above condition was confirmed by UV-Vis spectrophotometry, FT-IR, and NMR techniques.

***2.10.1. Spectrophotometric analysis.*** After cooling in icy bath to stop the reaction, without any extraction steps, the solution was analyzed by spectrophotometer using a Lambda Bio 20 spectrometer (Perkin-Elmer, Rotkreuz, Switzerland) using 1-cm absorption cell. Spectrum of MDA-PMA solutions was recorded from 200 to 600 nm at a scanning speed of one nm/min against of the glacial acetic acid as a blank reaction mixture. Absorbance reading was done at 404nm. Data acquisition and processing were carried out with the Perkin-Elmer UV Winlab software.

***2.10.2. FT-IR analysis.***The infrared (IR) spectra of the samples were recorded in the range 400–4000 cm−1 on a Fourier transform infrared (FTIR) spectrometer (model Perkin-Elmer, Rotkreuz, Switzerland) using KBr pellet method.

***2.10.3. Nuclear magnetic resonance spectroscopy.*** NMR spectra were performed on a 250 MHz Brüker (Germany) in deuterated chloroform for both experiments at room temperature to find whether the imine was formed or not.

**3. RESULTS AND DISCUSSION**

***3.1. Influence of time in derivative reaction***

The yellow complex under 30 oC cannot be produced, although time and concentration of Anisidine equal to 500 μM have been raised. However, at 90 and 75oC, reaction accelerates in short time (10min); this vital point can help us to consider the reaction of Imine formation or the same yellow complex in the real biological samples to determine MDA.

According to **Figure.3,** the temperature of 75 oC indicates the complete derivatization and the formation of the colorimetric reaction. Moreover, at 90oC, the reaction was carried out in a short time. However, at high level of temperature, the aldehyde levels may be raised in the real samples from the actual results which make it difficult to evaluate the data (Seljeskog et al.,2006). Therefore, to implement this method in biological samples, using 75 oC as an optimal temperature can be suited for the analysis. Furthermore, to obtain complete derivative reaction with Anisidine (500µM) at 75 oC, 20 min would be a proper time. Actually, by using 250 µM or lower concentrations of Anisidine, the complex (MDA-PMA) was remained at an intermediate state and the yellowish complex will not be formed and its result will not be suitable for the real samples. Therefore, the reaction is completed by Anisidine (500 μM) with optimized temperature at 75 oC within 20 min. (Figure 4.1 and Figure 4.2)



**Fig. 3.** Formation of yellow complex at different temperature (30,75, 90 oC)

**Fig.4.1.** The relationship between different anisidine concentration and time in complex (MDA-PMA) formation at 30 oC

**Fig. 4.2** The relationship between different anisidine concentration and time in complex (MDA-PMA) formation at 75 0C

On the other hand, TFEH, and PFH as derivative reagents require harsh temperature to produce complex (Giera *et al.,*2012 ).Temperature effect on derivative procedure was considered as an important part in this research; therefore, the samples placed in a water bath at 30, 75 and 90 oC and room temperature, then, cooled in icy bath.

To evaluate the optimum condition for formation of the complex [MDA-PMA], different concentrations of Anisidine (30, 50, 100, 150, 250, 500 µM) was added to different concentrations of MDA (3.125, 12.5 and 50 μM) in different times (5, 10, 20, 30 and 40) and in separate runs.

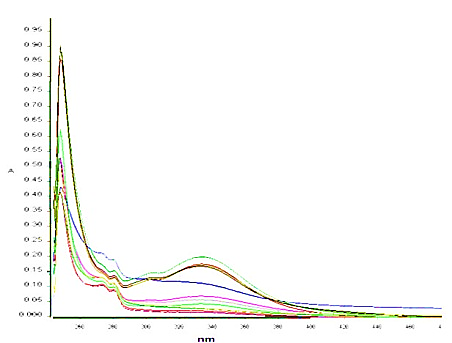
The results of Fig.4.1 indicate that yellow adduct was not formed at 30 oC and room temperature. Optimal temperature and time as the main part of derivation process were considered in this study. Under low temperature (30 oC), the reaction was slow and proceeded to produce an intermediate compound. Even, by increasing in the molar concentration of anisidine and time, it produces higher concentration of intermediate

compounds; this indicates that derivative adduct could not be formed in this concentration and temperature. However, while the concentration of Anisidine is 10 times higher than MDA concentration, the yellow color reduces the form of intermediate compounds and enhances the form of complex.

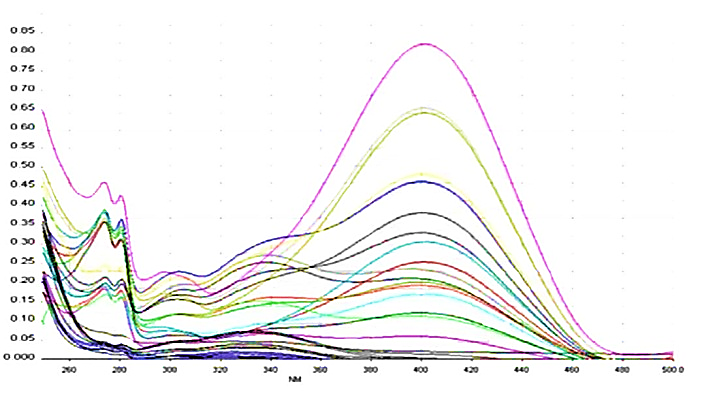
It seems that the procedure was completed at 90 oC in 10 min which contribute to form the imine reaction or yellow adduct in this temperature while this should be considered that high temperature is not proper for this procedure due to evaporation of malondialdehyde and solvents during derivative process. So this may be difficult to maintain the derivative concentrations.

On the other hand, at 30oC, there is no yellowish complex; therefore, it is not a proper temperature to assess the complex formation. Nevertheless, at 75oC, the reaction was completed in 40 min and the concentration of anisidine selected at 500 μm while Mao’s method reaction time was 3 hours with FMOC reagent (Mao *et al.,* 2006).

As can be seen in Figure 5, at the wavelength of 300-380 nm, the wide peak is related to intermediate formation whereas in Figure 6, relatively high peak in the range of 400-404 nm indicates reduced concentration of intermediate and formation of adduct (or the same considered complex).



**Fig.5.** Different concentration of anisidine with their absorbance spectra of derivative complex at 30 oC in complete reaction (Complete reaction)



**Fig.6.** Different concentration of anisidine with their absorbance spectra of derivative complex at 90 oC in complete reaction (Complete reaction)

The colorant has a strong absorbance peak in the visible range in 400 nm related to electronic transference π→ π\* (Figure.6). The absorbance peaks in UV region are related to electron transference of benzene rings (Mendes *et al.,* 2009). The obtained peak in visible range in the range of 300 to 395 is related to the formation of intermediates and the peak value decreases by high temperature. Likewise, by altering anisidine concentration, the adduct peak value increases.

**3.2.** *Optimal condition for derivative procedure*

The reaction of Anisidine (para - methoxy aniline) with aldehydic groups can be considered as an addition reaction. The yellowish complex emerged from releasing H2O molecules present a stable adduct. Investigating the reaction depended on MDA concentration indicated that the more molar concentration of anisidine, the more increase in the concentration of yellow complex (MDA-PMA). Derivatized adduct was evaluated with the molar ratios of 1/2:1 to 1:10 (MDA: Anisidine). **Figure.4.2** depicts when the molar ratio of anisidine concentration is (10[MDA]), the derivative solution attains to a higher level; thus, the derivatization was completed at the molar ratio of 10:1.

**3.3.** *Stability*

Current method is significantly different from Mao J, Zhang H, et al. (Mao et al., 2006). Their method was time consuming procedure (4 hour) at 50 oC to reach near completion reaction (Mao et al., 2006). To prevent false results, stability of malondialdehyde and anisidine as a crucial methodological aspect of this study were evaluated by UV-Vis spectrophotometry.

The absorbance reading of PMA (1000 µM and 500 µM) and MDA (stock solution (5000 µM) and its working standard solution (50 µM)) was done at 273 nm and 267 nm, respectively. It is clearly observed in **Figure.7.1** that the solution of PMA was stable if it is protected from light. Nevertheless, it would be better to prepare 500 µM concentration of PMA at the time of the test. **Figure.7.2** depicts the satisfactory results of 5000 µM concentration of MDA stock solution in terms of stability. However, working standard solution did not prove to be stable over 30-days period. Thus, it is only possible to use stock solution to prepare working standard solution at the time of test because MDA becomes degradation less than 24 hour.

On the other hand, FMOC-hydrazine and DNPH (as derivative reagents) increased the derivative reaction under mild acidic conditions and lower temperatures, because of having identical hydrazine groups, which are strong nucleophiles and can readily react with aldehydes. These conditions prevent undesired artificial aldehyde formation generated during the sample pretreatment. The derivative reagents such as DNPH need several Liquid-Liquid extractions due to the excess of unreached DNPH. Therefore, separation of the MDA-DNPH will be difficult and because of DNPH instability, purification of DNPH should be performed on a daily basis (Mao *et al.,* 2006; Shibamoto *et al.,*2006).

Consequently, based on above aforementioned results, this method did not require multiple extraction steps for anisidine reagent and thus, no residue or cloudiness was appeared during and after derivative procedure, which made this procedure preferable over the other methods. Furthermore, the yellowish adduct has no hazardous material if keep it for a long time in a laboratory. A feature which is crucial in developing a precise and accurate quantitative method is the stability of PMA. FMOC-Hydrazone (as a reagent) is only stable for 72 h at room temperature whereas PMA is stable for at least 1 month at room temperature if it is protected from light (Mao *et al.,* 2006; Shibamoto *et al.,*2006).

**Fig.7.1.** Evaluation of PMA stability

**Fig.7.2.** Evaluation of MDA stability

**3.4.** *The effect of pH*

The influence of pH in this procedure was examined. As can be seen in **Figure 8,** derivative complex at different pH (3.0, 7.4, 12.2) was formed after 10 min at 90 oC. The maximum yield was achieved at 400 nm in each pH. The results may contribute to apply this procedure for determination of free and total MDA in biological samples at different pH for further studies.

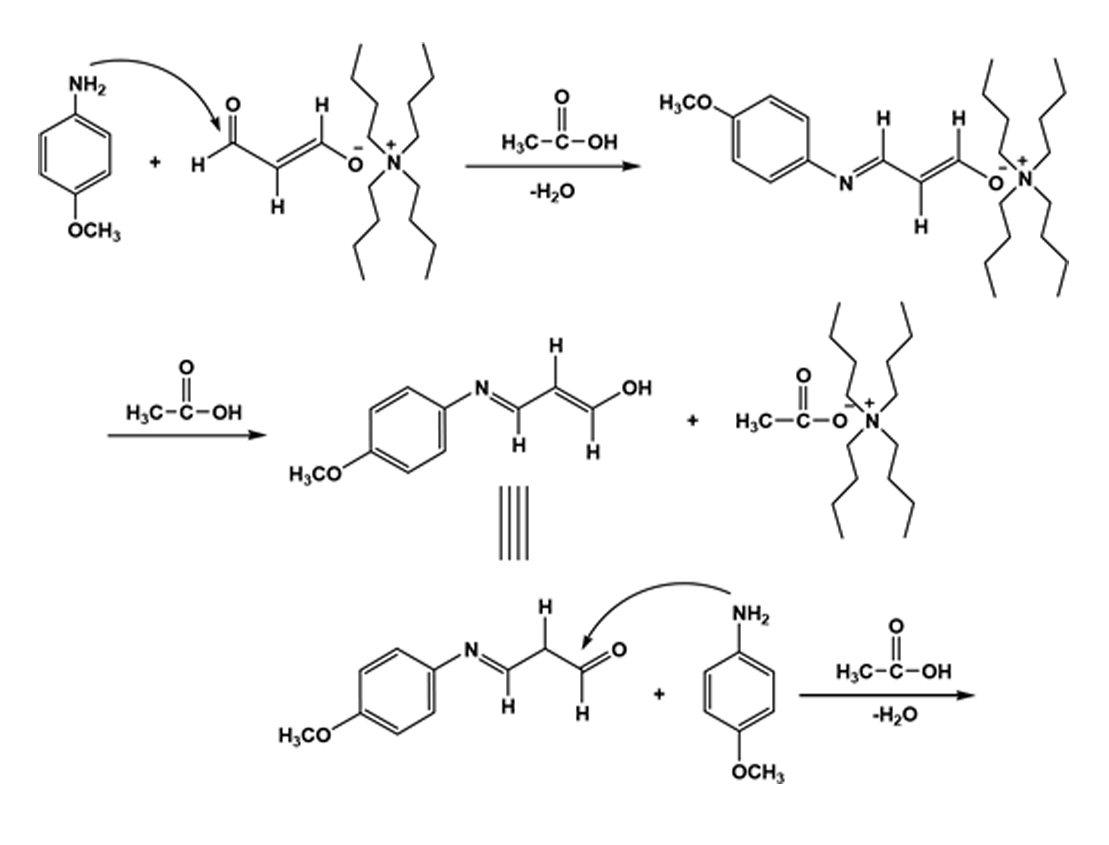
**Fig.8.** Evaluation of different pH for complex formation

**3.5.** *Mechanism of imine formation*

The formation of the imine is the initial step in this reaction. The imine was formed after reacting an amine (10) with aldehydes to form a dipolar intermediate. The reaction has two steps:

**1-**NH2 preferentially approach to carbonyl groups of aldehydes **2**- Elimination of water group.

In the current study, imine formed during the synthesis procedure identified by NMR and FT-IR techniques was confirmed as the adduct. The possible mechanism of MDA derivatization was depicted in **Figure.9**.



**N**

**H**

**3**

**CO**

**H**

**H**

**H**

**N**

**OCH**

**3**

**N**

**OCH**

**3**

**H**

**H**

**N**

**H**

**3**

**CO**

**H**

**H**

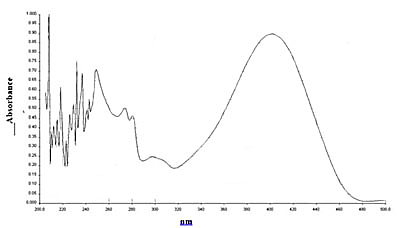
**H**

**Fig.9.**The mechanism of MDA-PMA adduct

**3.6.** *The results of UV-Vis, FT-IR, and NMR*

**3.6.1.** *Spectrophotometric results*

MDA and other carbonyl compounds naturally exist as byproducts of lipid peroxidation and prostaglandin biosynthesis. With regard to analytical approaches, it is better to derivatize MDA with reagents having high molar absorptivity at longer UV wavelengths (>254 nm). In fact, the high molar absorptivity and the close proximity of the absorbance maximum to ~ 300nm make an MDA derivative almost preferably suited for detection using a diode array system (Czauderna *et al.,* 2011). **(Figure.10)**



**Fig.10.** Spectrum of yellow adduct at 750C (50 µM MDA- 500 µM PMA)

**3.6.2.** *FT-IR results*

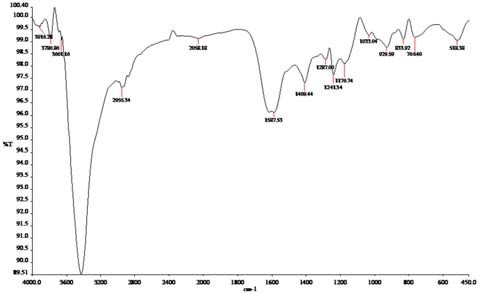
The FT-IR spectrums of MDA-derivatisation, primary Anisidine and malondialdehyde were confirmed in the reaction. IR spectra were different from spectra of MDA before and after derivative reaction. The important IR bands of the compounds along with their assignments are given in **Figures 11.1, 11.2, and 11.3.**



**NH2**

**OCH3**

**Fig.11.1.** IR-spectra of Anisidine



**H**

**O**

**O**

**H**

**H**

**N**

**Fig.11.2.** IR spectrum of Malondialdehyde tetrabutyl ammonium salt molecule



**N**

**H**

**3**

**CO**

**H**

**H**

**H**

**N**

**OCH**

**3**

**N**

**OCH**

**3**

**H**

**H**

**N**

**H**

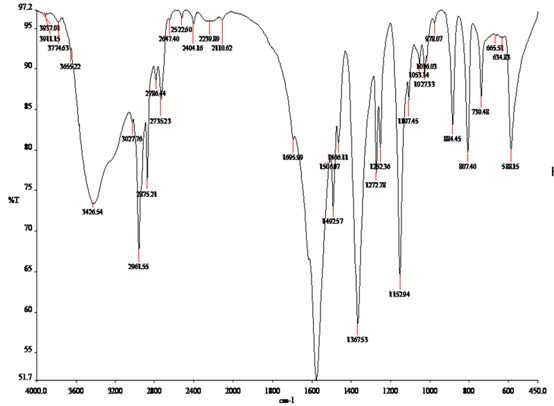
**3**

**CO**

**H**

**H**

**H**



**Fig.11.3.** IR spectrum of MDA-PMA and the proposed synthetic molecule

**Analytical data**

**Important infrared spectral (cm-1) bands of Anisidine**

Figure 11.1: IR-spectra of Anisidine demonstrate the N-H tension bond appears close 3346 and 3232cm-1. Another band at 3103 cm-1 was proved the aromatic CH tension. Furthermore, signals between 2838 cm-1 correspondent to the C-H aliphatic. Moreover, Band at 1510 shows the C=C and signals at 1334 and 1459 cm-1 depict the tension band of the CH3 and N-H, respectively. The stretching vibration of the C-N and C-O band for the methoxy group were appeared at 1235 cm-1 and 1030, respectively.

**Important infrared spectral (cm-1) bands of Malondialdhyde**

Figure 11.2: IR-spectra of MDA show the C-N stretching band (calculated at 1272 cm-1 ) related to the amine bands. The C-O stretching enol form band is clarified at 1152 cm-1 . The strong peak exhibits the aldehydic form of C=O at 1695 cm-1. The enol form of C=C was verified by the strong band at 1580 cm-1 . Besides, the CH3 and CH2 bending at 1367 and 1492 cm -1are roughly visible.

**IR Spectrum of Adduct**

Figure 11.3: IR-spectra of adduct shows Tension signals were eliminated at 2786 and 2735 cm-1 (C-H) due to the reaction between Anisidine and aldehydic group. Further band at 2955 cm-1 appeared as a result of *α,β*-unsaturated conjugated imides in <2955 cm -1 .

The new adduct being formed in acetic acid medium is conjugated imine aromatic. Not only is acetic acid as a catalyst for the reaction of amine and MDA but also make tetra-butyle amine separate from MDA and provide possible reaction for MDA and Anisidine again.

The important infrared spectral (cm-1) bands of MDA-PMA are considered in table.1 as follows:

**Table.1**.

**FT-IR results of MDA-PMA Adduct**

C-H = 2850 cm-1 N-H (NH group) = 3220 cm-1

C-N (amine group) = 1241 cm-1CH3 (bending vibration) = 1287cm-1

C-O (enole form) = 1176 cm-1 C=C (α, β unsaturated) = 1587 cm-1

C=N (imide) = 1605 cm-1 N-H = 1409 cm-1

Ammonium ion peak as a byproduct was appeared in a strong state at 3450 cm-1 .This is

the byproduct which has not been fully separated completely. **(Figure. 12)**

**N**

**H**

**3**

**C**

**C**

**O**

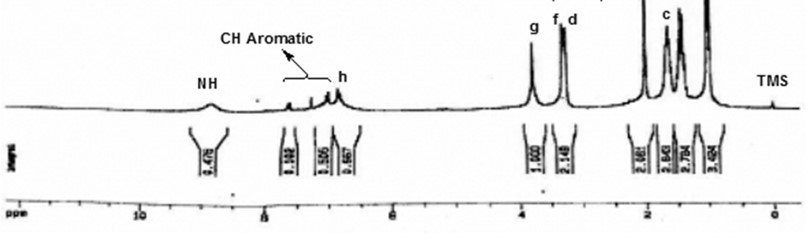
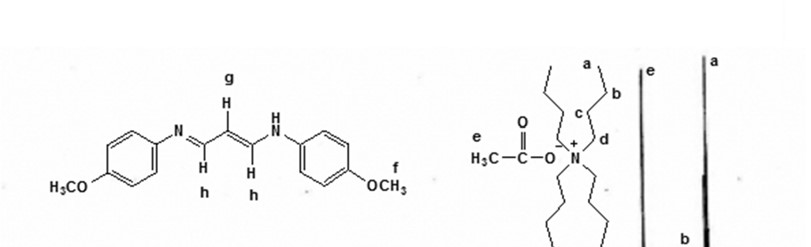
**O**

**Fig.12.** The byproduct of MDA-PMA reaction

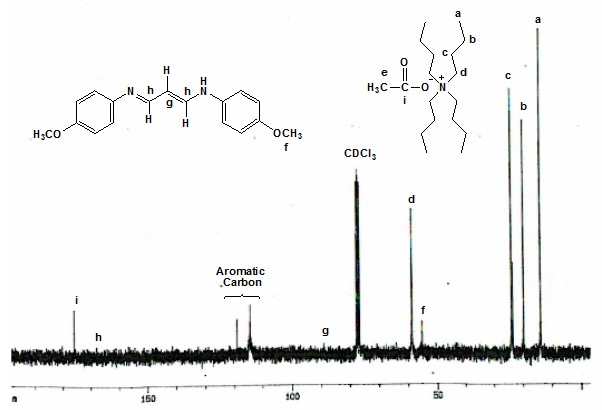
**3.6.3.** *NMR results*

The structure of adduct (MDA-PMA) is confirmed by the 1H-NMR and 13C-NMR. The protons of NH group in CDCL3 are shown in 9.8 ppm. **Figures 13.1, and 13.2.**

Fig. 10-3: Spectrum of adduct



**Fig.13.1.**1H NMR spectra of the adduct (MDA-PMA)



**Fig.13.2.**13C NMR spectra of main production with glacial acid acetic and tetra butyl ammonium salt

**Acknowledgements**

The authors would like to express their sincere appreciation to **Dr. Amirahmadi** for her suggestions during this research study.

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